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# Photochromism of dihydroindolizines. Part XV: Synthesis and photophysical properties of dihydroindolizine photoswitches bearing a conjugated aryleneethynylene tripodal linker system

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**Abstract** Different photochromic dihydroindolizines (DHIs) bearing conjugated aryleneethynylene tripodal linker systems were synthesized using several Sonogashira coupling reactions. The spirocyclopropene precursors incorporating different acetylenic bridge moieties at the 2-position of the fluorene moiety were synthesized via chemical and photochemical routes. Multiaddressable photochromic properties of the DHI derivatives substituted in the fluorene (region A) and pyridazine (region C) parts were studied. Optimization of the formation of the DHIs was also done by applying different palladium-mediated Sonogashira coupling reactions. Irradiation of the photochromic DHIs with polychromatic light led to colored betaines which undergo thermal 1,5-electrocyclization. The kinetics of the thermal 1,5-electrocyclization were studied by using a multichannel FT-UV-Vis spectrophotometer. A pronounced effect on the kinetic behavior of the 1,5-electrocyclization process of the betaines was observed by changing substitution from non-substituted to dimethyl-substituted pyridazines. Photodegradation experiments and the bleaching and fading cycles revealed high photostability of the betaines under investigation. These properties of betaines of tripodal linker conjugates will help these materials to find applications.

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#### Introduction

Photochromic molecules attract much attention both theoretically and practically because of their potential applications in optical devices, for example optical memories and switches [1-4]. These molecules find use in the fabrication of several optoelectronic devices, for example optical memories, switches, and holograms. Since the last decade, the development of such optoelectronic devices has included photochromic compounds as the active components [5]. It has been established that photochromic compounds support their activities in a reversible photochemical reaction induced by the absorption of electromagnetic radiation in the ultraviolet region, which provokes a visible color change of the original colorless molecule. Photochromic dihydroindolizines (DHIs), which were discovered and developed by H. Dürr [6–19], are well known photochromic materials that have attracted much interest from the standpoints of both fundamental elucidation of electrocyclization reactions and their potential applications to optical memories and switches [20-24, 38-44]. Because of a 1,5-electrocyclization photoisomerization between two distinct isomeric states, i.e., the ringopened form (betaine form) and the ring-closed form (DHI form), these entities are promising candidates for optical storage media and electronic devices [38-47]. Photochromic DHIs have sufficient thermal stability in both the open and the closed forms, very high resistance to photofatigue, and their chromophores provide convenient distinctive groupings that absorb ultraviolet radiation and allow the reaction to be easily monitored by UV–Vis spectroscopy [15–47].

Research on the synthesis of carbon-rich organic and organometallic compounds for widespread applications in the field of materials science has increased substantially. In this context, the use of  $\pi$ -conjugated rigid fluorenyl chromophores and their derivatives offers exciting perspectives for the design of new molecular oligomeric and polymeric materials for various optoelectronic applications [48–52]. Many molecular structures have been synthesized for use as molecular devices, including switches, wires, controllers, and gates [53–57]. Within contemporary acetylene chemistry [58], there is great interest in the synthesis of conjugated diyne and oligoyne molecules. These carbonrich building blocks [59] are fundamentally important for molecular rods and cyclic frameworks and are active components in optoelectronic devices such as wires, switches, and nonlinear optics, etc. [60–64].

In continuation of our research on the synthesis and photochromic behavior of photochromic DHIs, we now describe the synthesis and photophysical properties of carbon-rich fluorenyl DHI tripodal linkers and report studies on their photochromic behavior in solution. Also, synthesis approaches and Sonogashira-mediated methodologies are described in detail.

#### **Results and discussion**

# Synthesis of fluorenylethynylene spirocyclopropene precursors **8a–8c**

Dimethyl 2'-iodospiro[cycloprop[2]ene-1,9'-fluorene]-2,3dicarboxylate (1) has been previously prepared by us [65-67]. The spirocyclopropene precursors 8a (n = 0), 8b (n = 1), and **8c** (n = 2) were synthesized via a five-step sequence, starting with the known three-step conversion of 2-nitro-9H-fluoren-9-one to 2-iodo-9H-fluoren-9-one (2) in 64% yield [68, 69 and references therein] (Scheme 1). Sonogashira coupling of 2 with trimethylsilane derivatives **3a-3c** (n = 0, 1, 2) in the presence of Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub> (2.5%)/Cu<sub>2</sub>I<sub>2</sub>/Et<sub>2</sub>NH in THF for 4 h afforded the coupling products 4a (n = 0), 4b (n = 1), and 4c (n = 2) in 74, 67, and 59% yield, respectively. Condensation of the substituted fluorenones 4a-4c with hydrazine hydrate in boiling ethanol for 3 h led to both condensation and desilvlation with formation of **5a** (n = 0), **5b** (n = 1), and **5c** (n = 2)in 61, 54, and 42% yield, respectively.

The diazofluorene derivatives 6a-6c were obtained in 53–76% yields after oxidation of hydrazones 5a-5c with manganese dioxide in dry ether at room temperature in the absence of light. Addition of dimethyl acetylenedicarboxylate (DMAD) to 6a-6c in dry ether under dark conditions

for 8 h led to the formation of the pyrazole derivatives 7a-7c in moderate yields (55, 52, and 49%).

The target spirocyclopropene derivatives **8a–8c** were obtained in low yields ranging from 19 to 32% via photolysis of the pyrazole derivatives **7a–7c**. The photolysis was carried out in an inert and dry nitrogen atmosphere using a high pressure mercury lamp (125 W) in dry ether solution for 2 h. The chemical structures of the synthesized compounds **4–8** (Scheme 1) were confirmed and established by both spectroscopic (NMR, IR, and mass spectrometry) and elemental microanalytical data (elemental analysis data were satisfactory).

# Approaches to the synthesis of the $\pi$ -conjugated photochromic dihydroindolizines **10a–10f**

Electrophilic addition of spirocyclopropenes 8a-8c to pyridazine (9a) and 3,6-dimethylpyridazine (9b) using the cyclopropene route [11–47] (Scheme 2) under dry, dark, and inert conditions for 12 h (TLC monitored using CH<sub>2</sub>Cl<sub>2</sub> as eluent) led to the formation of the photochromic DHIs 11a-11f in poor yields (22-35%, method A). Formation of the photochromic DHIs 11a-11f occurs through conjugate addition of one of the nitrogen atoms of pyridazines 9a, 9b to of the electron-deficient  $\alpha,\beta$ -unsaturated spirocyclopropene diester 8a-8c, which leads to ring opening via a cyclopropyl-allyl conversion to the colored betaines 10a-10f (Scheme 2). A subsequent ring closure to DHIs 11a-11f results in a partial, slow, thermal 1,5-electrocyclization back reaction (Scheme 2) which can be reversed upon exposure to UV light. The photochromic DHIs 11a-11f were obtained in the pure form by purification on silica gel using dichloromethane as the eluent.

Alternatively, the target photochromic DHIs **11a–11f** could be synthesized via the multi-step synthesis route shown in Scheme 3. The palladium-mediated Sonogashira coupling of DHI **12a** and its dimethyl derivative **12b** [65–67] with compounds **3a–3c** (2.5% Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub>, CuI/Et<sub>3</sub>N, dry THF, 5 h) yielded the desired photochromic trimethylsilyl DHIs **13a–13f** through the betaines **14a–14f**, in 55–67% isolated yields. Purification was carried out by flash chromatography on silica gel with CH<sub>2</sub>Cl<sub>2</sub> as eluent (method B). Subsequent desilylation of the DHIs **13a–13f** with tetrabutylammonium fluoride (TBAF) in dry THF for 4 h afforded the silyl-free products **15a–15f** obtained in good isolated yields (62–75%).

The treatment of DHIs **13a–13f** with hydrazine hydrate in ethanol at 0 °C for 1 h afforded the desilylated photochromic DHIs **11a–11f** in 45–59% yield (method C), which is similar to results for compounds **4a–4c**. The products obtained from the three different routes gave the same analytical and spectroscopic data and the same melting points and mixed melting points.



COOCH,

#### Scheme 1

Palladium-mediated Sonogashira coupling of photochromic DHIs **11a–11f** with the tripodal linker **15** under different coupling conditions

The tripodal linker **15** was prepared in six steps and 29% overall yield in accordance with the methods reported by Galoppini and coworkers [70–80] and Zarwell and Rück-Braun [81]. The final fragment coupling of the photochromic DHIs **11a–11f** to the tripodal linker system **15** was surveyed,

via a Sonogashira cross-coupling reaction, under different reaction conditions for optimization purposes (Scheme 4). Reaction of **11a–11f** with **15** under standard Sonogashira coupling conditions  $(2\% \text{ Pd}(\text{PPh}_3)_2\text{Cl}_2, 1\% \text{ PPh}_3, \text{CuI/Et}_3\text{N}, \text{THF}, 15 \text{ h}$  at 45 °C, method A) led to low conversion of DHIs **11a–11f** to the coupling products **17a–17f** (22–28%). The conditions of the Sonogashira coupling were varied by using Pd(OAc)\_2/Ph\_3P with CuI/Et\_3N, toluene/DMF and stirring for 12 h at 40 °C (method B). In this case, the



Scheme 2



#### Scheme 3

coupling products **17a–17f** were obtained in slightly better yields (31–52%). With the objective of further improving the yields of **17a–17f**, the reaction conditions were modified to include bis(dibenzylideneacetone)palladium(0) as the new

palladium source, CuI, and DIEA as the base (method C), which led to the formation of the coupling products **17a–17f** in 52–68% yield. In addition, coupling of photochromic DHIs **11a–11f** with the tripod system **15** using 2.5 or 5%



#### Scheme 4

Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub>, 1% 1,10-phenanthroline, 5% Nal, CuI/Et<sub>3</sub>N, DMF at 90 °C (method D) did not afford the coupling products 17a-17f and, instead, decomposition of the reaction mixtures was observed.

Also the coupling reaction of DHIs **11a–11f** and the tripodal linker **15** using Pd(PPh<sub>3</sub>)<sub>4</sub> in dry pyrrolidine at 45 °C for 8 h (RT and 60 °C were also used) showed no evidence of the formation of the coupling products **17a–17f** (method E). This copper-free method was investigated to circumvent the homo-dimerization of the DHI system.

Attempts to improve the yield by manipulating the percentage of the palladium catalyst and reaction times were unsuccessful in all of the above mentioned coupling reactions. Interestingly, the photochromic DHIs **17a** and **17d** were previously synthesized by us [67] via a Sonogashira cross-coupling reaction by coupling of halo-substituted DHI with an acetylenic tripodal system which resulted in exactly the same chemical structures.

Attempted hydrolysis of the methyl ester groups of DHIs 17a-17f using KOH in ethanol or THF at room



Fig. 1 Representation of the optimized (MM2) structure of DHI 17c

temperature and at 50 °C afforded no product and instead decomposition of the DHIs was observed. All subsequent attempts to obtain the tricarboxylic acids 18a-18f by changing base concentration (4-8 M) and reaction times (from RT to 72 h, in ethanol or in THF) were met with failure. The chemical structures of all photochromic DHIs 11a-11f, 13a-13f, and 17a-17f were established on the basis of spectral and analytical data which corroborated the suggested chemical structures. For example, the chemical structure of DHI 17c was assigned by 2D NMR spectroscopy. Further assignments of 8'-CH, 8'a-CH, and some other protons in the DHI skeleton were achieved with the aid of a NOESY spectrum of 17c. We observed that 8'a-CH at  $\delta = 4.94$  ppm is proximal to 8'-CH at  $\delta = 5.35$  ppm and 1-CH of the fluorene moiety at  $\delta = 7.68$  ppm. This suggests that 8'a-CH is in the 8'a-position and not the 8'-position. Indeed, the connectivity between 8'-CH and 8-CH of the fluorene part at  $\delta = 7.54$  ppm was observed. This vicinity of 8'-CH with 8-CH indicates that the pyridazine moiety is perpendicular to the fluorene skeleton. This observation is also supported by molecular modeling calculation of DHI 17a (Fig. 1). The molecular mechanics calculation (MM2) showed that the distance between both 8'a-CH and 8'-CH connected to 6'-CH and 1-CH of the fluorene moiety is <3 Å, which is in good agreement with the NMR results (Fig. 1).

#### Absorption spectra of DHIs 11a–11f, 13a–13f, and 17a–17f, and their corresponding betaines 10a–10f, 14a–14f, and 16a–16f

The photophysical properties pertinent to their photochromic properties were obtained from the absorption features of photochromic DHIs **11a–11f**, **13a–13f**, and **17a–17f**. The absorption spectra of the synthesized DHIs **11a–11f**, **13a– 13f**, and **17a–17f** were measured in dichloromethane solution, at a concentration of  $1 \times 10^{-5}$  mol dm<sup>-3</sup>, at 23 °C, using a UV–Vis spectrophotometer. The photochromic DHIs 11a-11f. 13a-13f. and 17a-17f showed vellow color in dichloromethane solution and in the solid state (Table 1). The intensities (log  $\varepsilon$ ) of the absorption of these bands were found to lie between 3.80 and 4.67 depending on the number of aryleneethynylene groups. The absorption of DHIs 11a-11f, 13a–13f, and 17a–17f were observed in the far UV region and showed absorption maxima between 390 and 405 nm (Table 1). This absorption depends on the number of aryleneethynylene groups substituting the aromatic fluorene (region A). A pronounced bathchromic shift of about 6 nm was recorded after coupling of DHIs 11a-11f with the tripodal system 15, as in DHIs 17a-17f. This may be attributed to the increase in extended conjugation of the DHI skeleton following coupling. Exchanging the proton on the pyridazine with a methyl substituent led to a bathochromic shift of about 3 nm of the DHIs 17d-17f. These absorption bands can be assigned to the locally excited  $\pi - \pi^*$ -transition (LE) located in the butadienyl-vinylamine chromophores [11–47] of the photochromic DHIs 11a–11f, 13a–13f, and 17a-17f (Table 1).

Polychromatic light irradiation of DHIs 11a-11f, 13a-13f, and 17a–17f led to the ring-opened betaines 10a–10f, 14a-14f, and 16a-16f (Fig. 2). The colored betaine forms 10a-10f, 14a-14f, and 16a-16f changed from red to bluegreen in CH<sub>2</sub>Cl<sub>2</sub> solution, at a concentration of  $1 \times 10^{-5}$  mol dm<sup>-3</sup>, at room temperature because of their slower 1,5-electrocyclization. All absorption maxima of the colored betaines 10a-10f, 14a-14f, and 16a-16f were found in the visible region between 522 nm (betaine 11a) and 635 nm (betaine 17f). The UV spectra of the colored betaines containing a non-substituted pyridazine as a heterocyclic moiety, as in betaines 10a-10c, 14a-14c, and **16a–16c** (Fig. 2), exhibit a red-violet color and show only one absorption maximum ranging between 522 and 534 nm. On the other hand, the betaines containing a dimethylpyridazine in region C (10d-10f, 14d-14f, and 16d-16f) showed a green-blue color and three absorption maxima with three isobestic points (Table 1). Interestingly, a bathochromic shift by more than 110 nm was observed in the absorption spectra of the betaines containing nonsubstituted pyridazines 10a-10c, 14a-14c, and 16a-16c compared with the dimethyl analogs 10d-10f, 14d-14f, and 16d-16f. This large shift led to the change of the colored forms from red to green-blue, which is attributed to the hyper-conjugation bestowed by the two methyl groups which imparts stability to the zwitterionic betaines leading to a bathochromic shift [65–67] (Table 1). Furthermore, a noticeable bathochromic shift of about 7-9 nm was noted by increasing the number of bridged phenyl acetylenic groups from n = 0 to n = 2 which showed no dependence on the substitution pattern of the pyridazine. This may be attributed to an increase in the conjugation of the fluorene unit via hyperconjugation with the aromatic Table 1 Absorption spectra of DHIs 11a–11f, 13a–13f, and 17a–17f and their corresponding betaines 10a–10f, 14a–14f, and 16a–16f, and kinetic data of betaines 10a–10f, 14a–14f, and 16a–16f in the second range (monitored by UV spectrophotometry) in CH<sub>2</sub>Cl<sub>2</sub> solution (23 °C,  $c = 1 \times 10^{-5}$  mol dm<sup>-3</sup>)

DHI/Betaine	$\lambda_{\rm max}$ (DHI) (nm)	log (ɛ)	$\lambda_{\rm max}$ (betaine) (nm)	$k \times 10^{-3}$ (s)	<i>t</i> <sub>1/2</sub> (s)	Color of betaine
11a/10a	385	3.82	522	3.50	198	Red-violet
11b/10b	386	3.85	523	3.22	215	Red-violet
11c/10c	387	4.96	523	3.04	228	Red-violet
11d/10d	398	4.02	354, 444, 626	1.03	675	Green-blue
11e/10e	399	4.09	355, 445, 628	0.69	995	Green-blue
11f/10f	401	4.13	355, 448, 630	0.64	1,083	Green-blue
13a/14a	391	3.87	527	6.54	106	Red-violet
13b/14b	393	3.89	529	5.10	136	Red-violet
13c/14c	394	3.96	532	3.96	175	Red-violet
13d/14d	395	4.03	355, 449, 627	1.11	624	Red-violet
13e/14e	399	4.07	357, 445, 626	8.22	843	Green
13f/14f	402	4.13	352, 445, 634	7.40	937	Green
<b>17a/16a</b> <sup>a</sup>	402	4.22	529	2.57	270	Red-violet
17b/16b	403	4.28	532	2.42	287	Red-violet
17c/16c	405	4.35	534	2.31	300	Red-violet
17d/16d <sup>a</sup>	406	4.34	353, 442, 632	0.49	1,413	Green-blue
17e/16e	407	4.46	355, 447, 632	0.46	1,521	Green-blue
17f/16f	407	4.52	355, 448, 635	0.41	1,678	Green-blue

<sup>a</sup> The photochromic data of DHIs 17a, 17d have been reported elsewhere [65–67] and are cited here for comparison



Fig. 2 UV–Vis of photochromic DHI 11c and the corresponding betaine form 10c after UV irradiation in  $CH_2Cl_2(c = 1 \times 10^{-5} \text{ mol dm}^{-3})$  at ambient temperature

phenyl rings through the bridged acetylenic bond. Indeed, a bathochromic shift of about 10 nm of the absorption of the betaines **16a–16f** incorporating the tripod linker conjugate compared with the betaines **10a–10f** and **14a–14f** incorporating no tripodal linker conjugates was detected. More spectroscopic data about the UV–Vis measurements of the colored betaines under investigation are listed in Table 1.



**Fig. 3** FT–UV–Vis kinetic spectra of the 1,5-electrocylization of the betaine **14b** to DHI **13b** (cycle time = 50 s, run time = 400 s) in  $CH_2Cl_2$  ( $c = 1 \times 10^{-5}$  mol dm<sup>-3</sup> at 253 K)

The kinetics of the thermal 1,5-electrocyclization of the betaine forms 10a–10f, 14a–14f, and 16a–16f were studied by use of a multichannel FT–UV–Vis spectrophotometer (Figs. 3, 4). The thermal fading back reaction measurements showed that the half-lives of the colored betaines 10a–10f, 14a–14f, and 16a–16f lie in the second domain and were found to be between 106 and 1,678 s (Table 1



**Fig. 4** FT–UV–Vis kinetic spectra of the 1,5-electrocyclization of the betaine **16f** to DHI **17f** (cycle time = 500 s, run time = 4,000 s) in CH<sub>2</sub>Cl<sub>2</sub> ( $c = 1 \times 10^{-5}$  mol dm<sup>-3</sup> at 253 K)

and Figs. 3, 4). A pronounced increase in the half-lives of the betaines bearing a dimethylpyridazine 10d-10f, 14d-14f, and 16d–16f by approximately a factor of 7 compared with the half-lives of the betaines bearing a non-substituted pyridazine 10a-10c, 14a-14c, and 17a-17c was observed. As previously discussed [65-67], the detected increase in the half-lives may be attributed to the stabilization of the positive and negative charges on the betaine forms leading to enhancement of the half-lives of the betaine structures by the electron-donating methyl groups. Elongation of the half-lives of the betaine forms by increasing the number of acetylenic units in the bridge by approximately a factor of 1.30 was recorded. Also, a noticeable rise in the half-lives of the betaines coupled with tripodal linker system as in 16a–16f compared with non-coupled betaines 10a–10f and 14a-14f was observed. These increases and tuning of the absorption spectra and half-lives of the tripodal DHIs will help these materials to be supported on metal oxide nanoparticles.

#### Photo-degradation resistance of photochromic DHIs 11a–11f, 13a–13f, 17a–17f and their corresponding betaines 10a–10f, 14a–14f, 16a–16f

The gradual loss of the ability to change color by exposure to visible or ultraviolet light in this context has been termed fatigue [1–4, 11–14]. Gautron [82] has advanced a quantitative approach to measure fatigue in photochromic systems. Because of the slow thermal bleaching process of the betaines to DHIs, FT–UV–Vis measurement at room temperature was used to study the photo-fatigue of the DHIs and betaines under investigation. Irradiation of degassed dichloromethane solution of DHIs **11a–11f**, **13a**–

**Table 2** Photodegradation data of some selected betaines **10a–10f**, **14a–14f**, and **16a–16f** in dichloromethane solution ( $c = 1 \times 10^{-5} \text{ mol dm}^{-3}$ ) at 23 °C

Betaine	t <sub>30</sub> betaine/DHI	F	Betaine	t <sub>30</sub> betaine/DHI	F
10a	518	2.13	14d	498	2.05
10b	464	1.91	14e	451	1.86
10c	419	1.72	14f	436	1.79
10d	559	2.30	<b>16a</b> <sup>a</sup>	583	2.40
10e	520	2.14	16b	533	2.19
10f	499	2.05	16c	461	1.90
14a	461	1.90	<b>16d</b> <sup>a</sup>	694	2.86
14b	433	1.78	16e	599	2.47
14c	362	1.49	19f	512	2.11
Standard	243	1.00	Standard	243	1.00

<sup>a</sup> The photodegradation data of betaines **16a**, **16d** have been reported previously [65–67] and are cited here for comparison. The factor *F* is the ratio between the  $t_{30}$  value of the betaine under investigation and the standard betaine (dicyanopyridazine DHI)



**Fig. 5** Photodegradation experiment for determination of the  $t_{30^{-}}$  value of selected betaines **10a–10f**, **14a–14f**, and **16a–16f** in CH<sub>2</sub>Cl<sub>2</sub> ( $c = 1 \times 10^{-5} \text{ mol dm}^{-3}$ ) at ambient temperature

**13f**, **17a–17f** at room temperature (23 °C) with polychromatic light ( $\lambda = 200-400$  nm) led to the ring-opened colored betaines **10a–10f**, **14a–14f**, and **16a–16f**. Upon continued irradiation they decomposed after some time. The time in which the absorbance reaches 30% of its initial value is called  $t_{30}$  value.

The photodegradation data represented in Table 2 and Fig. 5 show that most of the selected betaines under investigation showed a higher photo-fatigue resistance than the standard dicyano-pyridazine DHI ( $t_{30} = 243$  min) by a factor between 1.72 as in the case of betaine **10c** and 2.86 as in the case of betaine **16d**. A noticeable decrease in the

 $t_{30}$  values by increasing the number of acetylenic bridges in both betaines bearing two methyl groups and the nonsubstituted pyridazines was recorded. A highly pronounced increase in the  $t_{30}$  values of the betaines incorporating dimethylpyridazine as in betaines 10d-10f, 14d-14f, and 16d-16f compared with the betaines incorporating a nonsubstituted pyridazine as in betaines 10a-10c, 14a-14c, and 16a-16c by factor of about 0.1-0.3 has been observed. Interestingly, betaines 16a–16f incorporating a tripodal linker showed the highest  $t_{30}$  values compared with other betaines under investigation. This interesting phenomenon will help these promising compounds find their applications. Betaine 16d ( $t_{30} = 694$  min) substituted with two methyl groups in the pyridazine moiety showed the highest photo-fatigue resistance amongst the other studied betaines and more than the standard betaine  $(t_{30} = 243 \text{ min})$  by a factor of 2.86.

The irradiation and thermal fading measurements presented in Fig. 6 show that the DHI/betaine **17e/16e** bearing the tripod linker system is more stable than the DHI/betaine **13e/14e** in terms of bleaching the thermal fading processes. The 20 bleaching and thermal fading cycles studied revealed no evidence of any decomposition product of the DHI/betaine **17e/16e**, as indicated by the UV–Vis measurements. These results have been further supported by NMR measurements which showed superimposable NMR spectra (coupling,  $\delta$  values, and intensities) of the DHIs before irradiation and after 20 cycles of irradiation with no detection of any decomposition products.

In conclusion, we have described different palladiummediated Sonogashira coupling reactions to optimize the coupling reactions with different acetylenic bridges and with the tripod linker system. Carbon-rich molecules and



Fig. 6 Irradiation and thermal fading of DHI/betaine 14e/13e and DHI/betaine 17e/16e (irradiation/thermal fading/irradiation cycles) in  $CH_2Cl_2$  ( $c = 1 \times 10^{-5}$  mol dm<sup>-3</sup>) at ambient temperature

tripodal linker conjugate based photochromic DHI derivatives were synthesized successfully through different pathways to optimize reaction conditions and yields. Many coupling reactions were done on the fluorene part (region A) with different phenyl acetylenic bridges to help the extension of the photochromism of target molecules for future applications in electronic devices, molecular wires, and solar energy conversion using different alternative routes. Interesting photochromic properties with tuning of the chemical structures of the photochromic DHIs by changing the number of acetylenic bridges in the fluorene part and substitutions in the pyridazine region were detected. The photodegradation measurements of the photochromic DHIs and their corresponding betaines under investigation showed a higher photo-fatigue resistance by direct irradiation or by cycling between colored and colorless forms than the standard dicyano-DHI system. The multi-addressable photochromic properties of the new studied photochromic DHIs and their corresponding betaines are expected to be helpful in finding suitable applications.

#### Experimental

Fluorenespirocyclopropene derivatives 8a-8c were obtained via photolysis of the corresponding pyrazoles in the photochemical reactor of Schenck [83] made from Pyrex  $(\lambda > 290 \text{ nm})$  according to reported procedures [15–53]. A high-pressure mercury lamp Philips-HPK (125 W) was used as a source of irradiation and the photolysis time was 4 h. Photochromic DHIs 17a and 17d were previously prepared by us [65–67]. The tripod linker system was prepared following the reaction procedures published by Galoppini and coworkers [70-80] and Zarwell and Rück-Braun [81]. Trimethylsilyl acetylene derivatives 2a-2c were prepared following the reaction procedures published by Rodriguez et al. [48-50, 68, 69]. Solutions to be photolyzed were flushed with dry nitrogen for 30 min before switching on the UV lamp. The progress of the reaction and the purity of the products isolated were monitored using TLC. Separation and purification of all synthesized photochromic materials were carried out using column chromatography  $(100 \times 2 \text{ cm})$  on silica gel and CH<sub>2</sub>Cl<sub>2</sub> as eluent. An Electrothermal Engineering meltingpoint apparatus was used to determine melting points. All NMR spectra were collected on a Bruker DRX 400 spectrometer (400 MHz) in CDCl<sub>3</sub> using TMS as the internal standard. Chemical shifts ( $\delta$ ) are reported in ppm. FT-IR measurements were performed using a Shimadzu FT-IR-8400S Fourier-transform infrared spectrophotometer. Mass spectra were recorded on a VG AutoSpec apparatus using electron impact at 70 eV. MALDI-MS spectra were

recorded in the positive mode using 2,5-dihydroxybenzoic acid in dioxane as matrix. UV-spectra were recorded on a Jasco V-570 FT-UV–Vis computerized spectrophotometer. All synthesized compounds gave satisfactory elemental analysis data. Compounds **17a** and **17d** are identical with those reported in Refs. [65–67].

General procedures for the synthesis of dimethyl 2-[ethynyl(phenylethynyl)<sub>n</sub>]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'dicarboxylates **11a–11f** 

Method A. A solution of spirocyclopropenes **8a–8c** (0.5 mmol) in 30 cm<sup>3</sup> dry ether and pyridazine (**9a**) or 3,6-dimethylpyridazine (**9b**) (0.7 mmol) were stirred at room temperature under dry N<sub>2</sub> in the absence of light for 12 h (TLC controlled). Ether was removed under reduced pressure and the products were purified by twice repeated column chromatography on silica gel using dichloromethane as eluent; finally recrystallization from the appropriate solvents afforded the products as pale yellow crystals.

Method B. To a solution of 2-iodopyridazine DHIs 12a or 12b (0.55 mmol) and trimethylsilylacetylene (3a), ((4ethynylphenyl)ethynyl)trimethylsilane (3b), or ((4-((4ethynylphenyl)ethynyl)phenyl)ethynyl)trimethylsilane (3c) (1.25 mmol) in 20 cm<sup>3</sup> dry THF and 50 cm<sup>3</sup> freshly distilled triethylamine under argon atmosphere at 50 °C was added dichlorobis(triphenylphosphine) palladium (19 mg, 0.026 mmol) and copper iodide (1.1 mg, 0.008 mmol). The mixture was stirred for 5 h, then the amine and THF were removed under reduced pressure. The crude residue was washed with saturated aqueous ammonium chloride solution  $(3 \times 20 \text{ cm}^3)$  with a small amount of KCN (200 mg), and extracted with dichloromethane  $(3 \times 20 \text{ cm}^3)$ . The extracts were dried over anhydrous sodium sulfate and the solvent was removed to give a brown solid, which was purified by column chromatography to afford the trimethylsilylated DHIs 13a-13f. The DHIs 13a-13f (0.14 mmol) were dissolved in 10 cm<sup>3</sup> freshly distilled dry THF and treated with a solution of tetrabutylammonium fluoride (TBAF, 0.150 g, 1.1 mmol) in 5 cm<sup>3</sup> freshly distilled THF at room temperature under nitrogen atmosphere. The reaction mixture was stirred for 4 h, then 10 cm<sup>3</sup> water was added. The mixture was extracted with ethyl acetate  $(3 \times 20 \text{ cm}^3)$ and the residue was purified by column chromatography using dichloromethane as eluent to afford the required compounds 11a-11f as yellow solids.

*Method C.* A solution of DHIs 13a-13f (0.25 mmol) and hydrazine hydrate (99%, 0.1 cm<sup>3</sup>, 2 mmol) in 10 cm<sup>3</sup> absolute ethanol was cooled to 0 °C and stirred at this temperature for 1 h. The solvent was removed, the product

was extracted three times with  $20 \text{ cm}^3$  ethyl acetate, and the residue was purified by column chromatography using dichloromethane as eluent to afford the required compounds **11a–11f** as yellow solids.

# $\label{eq:linear} \begin{array}{l} \textit{Dimethyl 2-ethynylspiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate} (\mathbf{11a}, \mathbf{C}_{25}\mathbf{H}_{18}\mathbf{N}_{2}\mathbf{O}_{4}) \end{array}$

Method A: yield 35%, m.p.: 177 °C; method B: yield 75%, m.p.: 177 °C; method C: yield 75%, m.p.: 176 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.91-7.92$  (d, J = 2.4 Hz, 1H, CH arom.), 7.87–7.89 (d, J = 2.4 Hz, 1H, CH arom.), 7.75-7.76 (d, J = 1.89 Hz, 1H, CH arom.), 7.49-7.52 (d, J = 2.4 Hz, 1H, CH arom.), 7.45–7.47 (d, J = 2.4 Hz, 1H, CH arom.), 7.40–7.42 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.30–7.33 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.21–7.23 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.72-5.74 (m, 1H, 7'-CH), 4.99-5.02 (t, J = 2.2 Hz, 1H, 8'-CH), 4.71–4.74 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 4.09 (s, 1H, acetylenic H), 3.87 (s, 3H, 3'-CH<sub>3</sub>), 3.45 (s, 3H, 2'-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.42$  (3'-CO), 161.79 (2'-CO), 149.23, 146.22, 142.97, 141.03, 140.19, 138.43, 136.02, 132.99, 130.62, 128.67, 126.76, 123.32, 122.46, 121.84, 120.34, 118.38, 104.48, 82.46 (acetylenic C), 81.34 (acetylenic C), 62.31 (8'a-C), 59.34 (spiro-C), 53.34  $(3'-CH_3)$ , 51.46  $(2'-CH_3)$  ppm; IR (KBr):  $\bar{v} = 3,076-$ 3,043 (C-H, arom.), 2,887-2,972 (C-H, aliph.), 2,162 (acetylenic bond), 1,741 (3'-C=O), 1,694 (2'-C=O), 1,585 (C=N), 1,432 (C=C), 1,357, 1,252, 1,179, 1,080, 953, 875, 760 cm<sup>-1</sup>; MS: m/z = 410.13 [M<sup>+</sup>].

### *Dimethyl* 2-[(4-ethynylphenyl)ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**11b**, C<sub>33</sub>H<sub>22</sub>N<sub>2</sub>O<sub>4</sub>)

Method A: yield 28%, m.p.: 164 °C; method B: yield 57%, m.p.: 165 °C; method C: yield 52%, m.p.: 163 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.88-7.90$  (d, J = 2.4 Hz, 1H, CH arom.), 7.87–7.89 (d, J = 2.4 Hz, 1H, CH arom.), 7.75–7.76 (d, J = 1.89 Hz, 1H, CH arom.), 7.56–7.58 (d, J = 7.98 Hz, 2H, CH arom.), 7.50–7.53 (d, J = 7.98 Hz, 2H, CH arom.), 7.45-7.47 (d, J = 2.4 Hz, 1H, CH arom.), 7.42–7.43 (d, J = 2.4 Hz, 1H, CH arom.), 7.39–7.41 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.29-7.32 (dt, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH)J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.23–7.25 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.70–5.73 (m, 1H, 7'-CH), 4.96–4.98 (t, J = 2.2 Hz, 1H, 8'a-CH), 4.70– 4.73 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 4.05 (s, 1H, acetylenic H), 3.82 (s, 3H, 3'-CH<sub>3</sub>), 3.42 (s, 3H, 2'-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.42$  (3'-CO), 161.79 (2'-CO), 149.23, 146.22, 142.97, 141.03, 140.19, 138.43, 136.02, 132.99, 131.56, 130.62, 128.67, 126.76, 123.32, 122.46, 121.84, 120.34, 118.38, 104.48, 93.12 (acetylenic C), 89.65 (acetylenic C), 82.46 (acetylenic C), 81.31 (acetylenic C), 62.30 (8'a-C), 59.35 (spiro-C), 53.39

(3'-CH<sub>3</sub>), 51.41 (2'-CH<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,100-3,025$  (C–H, arom.), 2,890–2,987 (C–H, aliph.), 2,154 (acetylenic bonds), 1,739 (3'-C=O), 1,692 (2'-C=O), 1,581 (C=N), 1,437 (C=C), 1,349, 1,254, 1,172, 1,081, 953, 877, 757 cm<sup>-1</sup>; MS: m/z = 510.16 [M<sup>+</sup>].

# Dimethyl 2-[[4-[(4-ethynylphenyl)ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**11c**, C<sub>41</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>)

Method A: yield 27%, m.p.: 144 °C; method B: yield 65%, m.p.: 143 °C; method C: yield 50%, m.p.: 143 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.85-7.86$  (d, J = 2.4 Hz, 1H, CH arom.), 7.86–7.88 (d, J = 2.4 Hz, 1H, CH arom.), 7.74–7.75 (d, J = 1.89 Hz, 1H, CH arom.), 7.56–7.58 (m, 6H, CH arom.), 7.50–7.53 (d, J = 7.98 Hz, 2H, CH arom.), 7.44–7.46 (d, J = 2.4 Hz, 1H, CH arom.), 7.40–7.42 (d, J = 2.4 Hz, 1H, CH arom.), 7.36–7.38 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.30–7.33 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.24–7.25 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.71–5.74 (m, 1H, 7'-CH), 4.95–4.97 (t, J = 2.2 Hz, 1H, 8'a-CH), 4.75–4.78 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 4.07 (s, 1H, acetylenic H), 3.84 (s, 3H, 3'-CH<sub>3</sub>), 3.45 (s, 3H, 2'-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.36$  (3'-CO), 161.85 (2'-CO), 149.13, 146.01, 143.10, 141.63, 140.23, 138.68, 136.00, 132.95, 131.95, 131.51, 130.69, 128.60, 126.79, 123.30, 122.44, 121.91, 120.37, 118.36, 104.54, 93.23 (acetylenic C), 89.77 (acetylenic C), 82.47 (acetylenic C), 81.43 (acetylenic C), 62.37 (8'a-C), 59.36 (spiro-C), 53.43  $(3'-CH_3)$ , 51.46  $(2'-CH_3)$  ppm; IR (KBr):  $\bar{v} = 3,100-3,019$ (C-H, arom.), 2,890-2,982 (C-H, aliph.), 2,157 (acetylenic bonds), 1,749 (3'-C=O), 1,694 (2'-C=O), 1,576 (C=N), 1,476 (C=C), 1,353, 1,257, 1,136, 1,088, 967, 871, 763 cm<sup>-1</sup>; MS: m/z = 610.19 [M<sup>+</sup>].

# Dimethyl (4'aS)-2-ethynyl-2',4'a-dimethylspiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**11d**, $C_{27}H_{22}N_2O_4$ )

Method A: yield 26%, m.p.: 146 °C; method B: yield 63%, m.p.: 145 °C; method C: yield 54%, m.p.: 144 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.95-7.97$  (d, J = 2.4 Hz, 1H, CH arom.), 7.84–7.86 (d, J = 2.4 Hz, 1H, CH arom.), 7.73–7.75 (d, J = 1.89 Hz, 1H, CH arom.), 7.50–7.54 (d, J = 2.4 Hz, 1H, CH arom.), 7.47–7.49 (d, J = 2.4 Hz, 1H, CH arom.), 7.37–7.40 (dt, J = 2.4 Hz, J = 0.88Hz, 1H, CH arom.), 7.18-7.22 (dt, J = 2.4 Hz, J =0.88 Hz, 1H, CH arom.), 5.60-5.63 (m, 1H, 7'-CH), 4.78–4.81 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 4.06 (s, 1H, acetylenic H), 3.84 (s, 3H, 3'-CH<sub>3</sub>), 3.44 (s, 3H, 2'-CH<sub>3</sub>), 2.11 (s, 3H, 6'-CH<sub>3</sub>), 1.43 (s, 3H, 8'a-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 164.00 (3'-CO), 161.98 (2'-$ CO), 149.26, 146.46, 143.89, 141.64, 140.19, 138.76, 136.36, 133.06, 130.66, 128.46, 126.76, 123.47, 122.63, 121.88, 120.42, 118.37, 104.52, 82.42 (acetylenic C), 81.36 (acetylenic C), 63.26 (8'a-C), 59.31 (spiro-C), 53.39 (3'-CH<sub>3</sub>), 51.25 (2'-CH<sub>3</sub>), 22.46 (6'-CH<sub>3</sub>), 21.22 (8'-CH<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,091-3,013$  (C–H, arom.), 2,846–2,979 (C–H, aliph.), 2,167 (acetylenic bond), 1,747 (3'-C=O), 1,693 (2'-C=O), 1,585 (C=N), 1,438 (C=C), 1,363, 1,250, 1,187, 1,082, 950, 878, 766 cm<sup>-1</sup>; MS: m/z = 438.16 [M<sup>+</sup>].

#### Dimethyl (4'aS)-2-[(4-ethynylphenyl)ethynyl]-2',4'a-dimethylspiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**11e**, C<sub>35</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>)

Method A: yield 24%, m.p.: 133 °C; method B: yield 68%, m.p.: 134 °C; method C: yield 49%, m.p.: 133 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.85-7.87$  (d, J = 2.4 Hz, 1H, CH arom.), 7.82-7.81 (d, J = 2.4 Hz, 1H, CH arom.), 7.71–7.74 (d, J = 1.89 Hz, 1H, CH arom.), 7.57–7.61 (d, J = 7.98 Hz, 2H, CH arom.), 7.54–7.56 (d, J = 7.98 Hz, 2H, CH arom.), 7.44–7.46 (d, J = 2.4 Hz, 1H, CH arom.), 7.40–7.43 (d, J = 2.4 Hz, 1H, CH arom.), 7.30–7.34 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.24–7.26 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.76–5.80 (m, 1H, 7'-CH), 4.74–4.77 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 4.04 (s, 1H, acetylenic H), 3.80 (s, 3H, 3'-CH<sub>3</sub>), 3.47 (s, 3H, 2'-CH<sub>3</sub>), 2.14 (s, 3H, 6'-CH<sub>3</sub>), 1.42 (s, 3H, 8'a-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.58$  (3'-CO), 161.92 (2'-CO), 149.26, 146.20, 143.03, 141.13, 140.34, 138.45, 136.12, 132.87, 131.54, 130.56, 128.64, 126.71, 123.31, 122.44, 121.83, 120.34, 118.33, 104.44, 93.10 (acetylenic C), 89.64 (acetylenic C), 82.44 (acetylenic C), 81.37 (acetylenic C), 62.32 (8'a-C), 59.37 (spiro-C), 53.30 (3'-CH<sub>3</sub>), 51.47 (2'-CH<sub>3</sub>), 22.45 (6'-CH<sub>3</sub>), 21.29 (8'a-CH<sub>3</sub>) ppm; IR (KBr):  $\bar{v} = 3,096-3,012$  (C–H, arom.), 2,880– 2,981 (C-H, aliph.), 2,150 (acetylenic bonds), 1,743 (3'-C=O), 1,690 (2'-C=O), 1,582 (C=N), 1,440 (C=C), 1,352, 1,250, 1,167, 1,089, 950, 889, 765 cm<sup>-1</sup>; MS:  $m/z = 538.19 [M^+].$ 

Dimethyl (4'aS)-2-[[4-[(4-ethynylphenyl)ethynyl]phenyl]ethynyl]-2',4'a-dimethylspiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate

 $(11f, C_{43}H_{30}N_2O_4)$ 

Method A: yield 22%, m.p.: 121 °C; method B: yield 62%, m.p.: 120 °C; method C: yield 45%, m.p.: 120 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.88–7.91 (d, *J* = 2.4 Hz, 1H, CH arom.), 7.85–7.87 (d, *J* = 2.4 Hz, 1H, CH arom.), 7.76–7.78 (d, *J* = 1.89 Hz, 1H, CH arom.), 7.54–7.59 (m, 6H, CH arom.), 7.51–7.53 (d, *J* = 7.98 Hz, 2H, CH arom.), 7.42–7.45 (d, *J* = 2.4 Hz, 1H, CH arom.), 7.38–7.41 (d, *J* = 2.4 Hz, 1H, CH arom.), 7.32–7.35 (dt, *J* = 2.4 Hz, *J* = 0.88 Hz, 1H, CH arom.), 7.20–7.24 (dt, *J* = 2.4 Hz, *J* = 0.88 Hz, 1H, CH arom.), 5.78–5.82 (m, 1H, 7'-CH), 4.72–4.76 (dt, *J* = 9.60 Hz, *J* = 1.76 Hz, 8'-CH), 4.04 (s, 1H, acetylenic H), 3.82 (s, 3H, 3'-CH<sub>3</sub>), 3.44 (s, 3H, 2'-CH<sub>3</sub>), 2.18 (s, 3H, 6'-CH<sub>3</sub>), 1.44 (s, 3H, 8'a-CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 163.45 (3'-CO), 161.84 (2'- CO), 149.13, 146.09, 143.18, 141.62, 140.18, 138.67, 136.05, 132.97, 131.98, 131.50, 130.73, 128.69, 126.83, 123.37, 122.42, 121.87, 120.43, 118.38, 104.53, 93.22 (acetylenic C), 89.74 (acetylenic C), 82.42 (acetylenic C), 81.40 (acetylenic C), 62.34 (8'a-C), 59.43 (spiro-C), 53.48 (3'-CH<sub>3</sub>), 51.46 (2'-CH<sub>3</sub>), 22.40 (6'-CH<sub>3</sub>), 21.34 (8'a-CH<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,089-3,023$  (C–H, arom.), 2,898-2,983 (C–H, aliph.), 2,152 (acetylenic bonds), 1,743 (3'-C=O), 1,689 (2'-C=O), 1,571 (C=N), 1,473 (C=C), 1,355, 1,253, 1,142, 1,067, 962, 885, 764 cm<sup>-1</sup>; MS: m/z = 638.22 [M<sup>+</sup>].

#### Dimethyl 2-[(trimethylsilyl)ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**13a**, C<sub>28</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>Si)

Yield 55%, m.p.: 121 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.87-7.90$  (d, J = 2.4 Hz, 1H, CH arom.), 7.83-7.86 (d, J = 2.4 Hz, 1H, CH arom.), 7.73-7.75 (d, J = 1.89 Hz, 1.89 Hz)1H, CH arom.), 7.44–7.46 (d, J = 2.4 Hz, 1H, CH arom.), 7.40–7.43 (d, J = 2.4 Hz, 1H, CH arom.), 7.36–7.39 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.32–7.35 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.18–7.22 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.69–5.72 (m, 1H, 7'-CH), 4.94–4.96 (t, J = 2.2 Hz, 1H, 8'a-CH), 4.70–4.73 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.86 (s, 3H, 3'-CH<sub>3</sub>), 3.47 (s, 3H, 2'-CH<sub>3</sub>), 0.94 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.37$  (3'-CO), 161.85 (2'-CO), 149.20, 146.36, 142.87, 141.26, 140.37, 138.47, 136.36, 132.90, 130.97, 128.64, 126.78, 123.32, 122.43, 121.58, 120.36, 118.21, 104.78, 82.37 (acetylenic C), 81.30 (acetylenic C), 62.46 (8'a-C), 59.57 (spiro-C), 53.30 (3'-CH<sub>3</sub>), 51.27 (2'-CH<sub>3</sub>), 13.85 (Si(CH<sub>3</sub>)<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,098-3,028$  (C–H, arom.), 2,880–2,987 (C–H, aliph.), 2,149 (acetylenic bond), 1,745 (3'-C=O), 1,689 (2'-C=O), 1,578 (C=N), 1,424 (C=C), 1,363, 1,254, 1,182, 1,036, 964, 870, 767 cm<sup>-1</sup>; MS: m/z = 482.17 [M<sup>+</sup>].

### Dimethyl 2-[[4-[(trimethylsilyl)ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**13b**, C<sub>36</sub>H<sub>30</sub>N<sub>2</sub>O<sub>4</sub>Si)

Yield 58%, m.p.: 149 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.95-7.98$  (d, J = 2.4 Hz, 1H, CH arom.), 7.88–7.90 (d, J = 2.4 Hz, 1H, CH arom.), 7.77–7.79 (d, J = 1.89 Hz, 1H, CH arom.), 7.52–7.55 (d, J = 7.98 Hz, 2H, CH arom.), 7.49–7.51 (d, J = 7.98 Hz, 2H, CH arom.), 7.44–7.46 (d, J = 2.4 Hz, 1H, CH arom.), 7.43–7.45 (d, J = 2.4 Hz, 1H, CH arom.), 7.38–7.42 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.31–7.34 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.20–7.24 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.76–5.79 (m, 1H, 7'-CH), 4.93–4.96 (t, J = 2.2 Hz, 1H, 8'a-CH), 4.68–4.72 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.81 (s, 3H, 3'-CH<sub>3</sub>), 3.46 (s, 3H, 2'-CH<sub>3</sub>), 0.91 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.65$  (3'-CO), 161.68 (2'-CO), 149.20, 146.37, 142.85, 141.11, 140.23, 138.47, 136.08, 132.87, 131.50, 130.76, 128.64, 126.64, 123.37, 122.54, 121.97, 120.30, 118.57, 104.36, 93.04 (acetylenic C), 89.53 (acetylenic C), 82.38 (acetylenic C), 81.37 (acetylenic C), 62.28 (8'a-C), 59.37 (spiro-C), 53.46 (3'-CH<sub>3</sub>), 51.74 (2'-CH<sub>3</sub>), 13.79 (Si(CH<sub>3</sub>)<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,098-3,029$  (C–H, arom.), 2,890–2,978 (C–H, aliph.), 2,155 (acetylenic bonds), 1,745 (3'-C=O), 1,689 (2'-C=O), 1,580 (C=N), 1,441 (C=C), 1,356, 1,257, 1,178, 1,064, 950, 887, 751 cm<sup>-1</sup>; MS: m/z = 582.20 [M<sup>+</sup>].

Dimethyl 2-[[4-[[4-[(trimethylsilyl)ethynyl]phenyl]ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2b]pyridazine]-6',7'-dicarboxylate (**13c**,  $C_{44}H_{34}N_2O_4Si$ )

Yield 61%, m.p.: 163 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.84-7.87$  (d, J = 2.4 Hz, 1H, CH arom.), 7.81-7.84 (d, J = 2.4 Hz, 1H, CH arom.), 7.70-7.74 (d, J = 1.89 Hz,1H, CH arom.), 7.51-7.54 (m, 6H, CH arom.), 7.46-7.49 (d, J = 7.98 Hz, 2H, CH arom.), 7.43-7.45 (d, J = 2.4 Hz,1H, CH arom.), 7.40–7.42 (d, J = 2.4 Hz, 1H, CH arom.), 7.35–7.37 (dd, J = 1.76 Hz, J = 1.98 Hz, 1H, 6'-CH), 7.32–7.35 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.26–7.28 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.70–5.73 (m, 1H, 7'-CH), 4.98–5.01 (t, J = 2.2 Hz, 1H, 8'a-CH), 4.73–4.75 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.80 (s, 3H, 3'-CH<sub>3</sub>), 3.45 (s, 3H, 2'-CH<sub>3</sub>), 0.89 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.45$ (3'-CO), 161.88 (2'-CO), 149.12, 146.34, 143.18, 141.57, 140.20, 138.76, 136.08, 132.79, 131.35, 131.50, 130.76, 128.58, 126.84, 123.36, 122.47, 121.87, 120.36, 118.34, 104.53, 93.64 (acetylenic C), 89.76 (acetylenic C), 82.23 (acetylenic C), 81.10 (acetylenic C), 62.27 (8'a-C), 59.31 (spiro-C), 53.47 (3'-CH<sub>3</sub>), 51.52 (2'-CH<sub>3</sub>), 13.81  $(Si(CH_3)_3)$  ppm; IR (KBr):  $\bar{v} = 3,087-3,021$  (C–H, arom.), 2,894-2,978 (C-H, aliph.), 2,152 (acetylenic bonds), 1,745 (3'-C=O), 1,691 (2'-C=O), 1,573 (C=N), 1,477 (C=C), 1,352, 1,254, 1,122, 1,046, 960, 877, 763 cm<sup>-1</sup>; MS:  $m/z = 682.23 \, [M^+].$ 

#### *Dimethyl* (4'aS)-2',4'a-dimethyl-2-[(trimethylsilyl)ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**13d**, C<sub>30</sub>H<sub>30</sub>N<sub>2</sub>O<sub>4</sub>Si)

Yield 67%, m.p.: 112 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.96-7.99$  (d, J = 2.4 Hz, 1H, CH arom.), 7.88–7.92 (d, J = 2.4 Hz, 1H, CH arom.), 7.76–7.78 (d, J = 1.89 Hz, 1H, CH arom.), 7.54–7.53 (d, J = 2.4 Hz, 1H, CH arom.), 7.44–7.47 (d, J = 2.4 Hz, 1H, CH arom.), 7.39–7.42 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.20–7.24 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.72–5.74 (m, 1H, 7'-CH), 4.80–4.83 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.84 (s, 3H, 3'-CH<sub>3</sub>), 3.44 (s, 3H, 2'-CH<sub>3</sub>), 2.11 (s, 3H, 6'-CH<sub>3</sub>), 1.43 (s, 3H, 8'a-CH<sub>3</sub>), 0.90 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.89$  (3'-CO), 161.94 (2'-CO), 149.30, 146.21, 143.85, 141.63, 140.27,

138.26, 136.33, 133.09, 130.57, 128.42, 126.61, 123.46, 122.60, 121.74, 120.39, 118.46, 104.57, 82.41 (acetylenic C), 81.22 (acetylenic C), 63.32 (8'a-C), 59.21 (spiro-C), 53.46 (3'-CH<sub>3</sub>), 51.23 (2'-CH<sub>3</sub>), 22.41 (6'-CH<sub>3</sub>), 21.28 (8'-CH<sub>3</sub>), 13.86 (Si(CH<sub>3</sub>)<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,098-3,008$  (C-H, arom.), 2,842–2,976 (C-H, aliph.), 2,165 (acetylenic bond), 1,745 (3'-C=O), 1,685 (2'-C=O), 1,581 (C=N), 1,442 (C=C), 1,359, 1,257, 1,178, 1,081, 958, 877, 763 cm<sup>-1</sup>; MS: m/z = 510.20 [M<sup>+</sup>].

### Dimethyl (4'aS)-2',4'a-dimethyl-2-[[4-[(trimethylsilyl)ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**13e**, C<sub>38</sub>H<sub>34</sub>N<sub>2</sub>O<sub>4</sub>Si)

Yield 64%, m.p.: 125 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.84-7.86$  (d, J = 2.4 Hz, 1H, CH arom.), 7.80-7.83 (d, J = 2.4 Hz, 1H, CH arom.), 7.75-7.79 (d, J = 1.89 Hz,1H, CH arom.), 7.52-7.55 (d, J = 7.98 Hz, 2H, CH arom.), 7.48–7.51 (d, J = 7.98 Hz, 2H, CH arom.), 7.40–7.44 (d, J = 2.4 Hz, 1H, CH arom.), 7.36–7.39 (d, J = 2.4 Hz, 1H, CH arom.), 7.32-7.37 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 7.21–7.23 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.69-5.74 (m, 1H, 7'-CH), 4.71-4.74 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.84 (s, 3H, 3'-CH<sub>3</sub>), 3.42 (s, 3H, 2'-CH<sub>3</sub>), 2.11 (s, 3H, 6'-CH<sub>3</sub>), 1.47 (s, 3H, 8'a-CH<sub>3</sub>), 0.87 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.46$  (3'-CO), 161.87 (2'-CO), 149.32, 146.28, 143.46, 141.78, 140.36, 138.41, 136.10, 132.74, 131.68, 130.26, 128.64, 126.77, 123.26, 122.47, 121.82, 120.27, 118.46, 104.48, 93.11 (acetylenic C), 89.54 (acetylenic C), 82.47 (acetylenic C), 81.36 (acetylenic C), 62.29 (8'a-C), 59.37 (spiro-C), 53.28 (3'-CH<sub>3</sub>), 51.47 (2'-CH<sub>3</sub>), 22.46 (6'-CH<sub>3</sub>), 21.31 (8'a-CH<sub>3</sub>), 13.78 (Si(CH<sub>3</sub>)<sub>3</sub>) ppm; IR (KBr):  $\bar{v} = 3,110-3,043$  (C–H, arom.), 2,895– 2,981 (C-H, aliph.), 2,152 (acetylenic bonds), 1,741 (3'-C=O), 1,692 (2'-C=O), 1,588 (C=N), 1,446 (C=C), 1,375, 1,263, 1,146, 1,087, 955, 896, 761 cm<sup>-1</sup>; MS:  $m/z = 610.23 \, [M^+].$ 

### Dimethyl (4'aS)-2',4'a-dimethyl-2-[[4-[[4-[(trimethylsilyl)ethynyl]phenyl]ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**13f**, C<sub>46</sub>H<sub>38</sub>N<sub>2</sub>O<sub>4</sub>Si)

Yield 61%, m.p.: 143 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 7.98-8.03$  (d, J = 2.4 Hz, 1H, CH arom.), 7.92–7.94 (d, J = 2.4 Hz, 1H, CH arom.), 7.81–7.85 (d, J = 1.89 Hz, 1H, CH arom.), 7.50–7.55 (m, 6H, CH arom.), 7.46–7.50 (d, J = 7.98 Hz, 2H, CH arom.), 7.40–7.43 (d, J = 2.4 Hz, 1H, CH arom.), 7.26–7.30 (d, J = 2.4 Hz, 1H, CH arom.), 7.26–7.30 (d, J = 2.4 Hz, 1H, CH arom.), 7.26–7.41 (dt, J = 2.4 Hz, 1H, CH arom.), 7.16–7.19 (dt, J = 2.4 Hz, J = 0.88 Hz, 1H, CH arom.), 5.85–5.49 (m, 1H, 7'-CH), 4.70–4.74 (dt, J = 9.60 Hz, J = 1.76 Hz, 8'-CH), 3.87 (s, 3H, 3'-CH<sub>3</sub>),

3.44 (s, 3H, 2'-CH<sub>3</sub>), 2.13 (s, 3H, 6'-CH<sub>3</sub>), 1.42 (s, 3H, 8'a-CH<sub>3</sub>), 0.86 (s, 9H, Si(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 163.46$  (3'-CO), 161.72 (2'-CO), 149.27, 146.16, 143.36, 141.84, 140.28, 138.60, 136.16, 133.13, 131.85, 131.49, 130.73, 128.67, 126.75, 123.32, 122.40, 121.79, 120.52, 118.46, 104.79, 93.29 (acetylenic C), 89.58 (acetylenic C), 82.46 (acetylenic C), 81.42 (acetylenic C), 62.79 (8'a-C), 59.41 (spiro-C), 53.36 (3'-CH<sub>3</sub>), 51.54 (2'-CH<sub>3</sub>), 22.28 (6'-CH<sub>3</sub>), 21.34 (8'a-CH<sub>3</sub>), 13.81 (Si(CH<sub>3</sub>)<sub>3</sub>) ppm; IR (KBr):  $\bar{\nu} = 3,085-3,013$  (C–H, arom.), 2,898–2,967 (C–H, aliph.), 2,145 (acetylenic bonds), 1,746 (3'-C=O), 1,678 (2'-C=O), 1,564 (C=N), 1,458 (C=C), 1,351, 1,247, 1,136, 1,067, 960, 879, 763 cm<sup>-1</sup>; MS: *m*/*z* = 710.26 [M<sup>+</sup>].

# General procedure for the synthesis of photochromic DHIs-tripodal linker 17b, 17c, 17e, and 17f

Method A. To an oven-dried screw-cap tube or a roundbottomed flask equipped with a water cooled West condenser and a magnetic stir bar the 2-iodosubstituted fluorene DHIs 11b, 11c, 11e, and 11f (3 mmol), 20 mg palladium di(triphenylphosphine) dichloride (0.06 mmol), 78 mg triphenylphosphine (0.03 mmol), and 36 mg CuI (0.15 mmol) were added to 1-(4-iodophenyl)-3,5,7-tris[4-(ethoxycarbonyl)phenyl]adamantane **15** (3 mmol). The vessel was then sealed with a rubber septum, evacuated, and backfilled with nitrogen (three times). A co-solvent system of 20 cm<sup>3</sup> freshly distilled THF and 0.83 cm<sup>3</sup> triethylamine (6 mmol) was added. The reaction mixture was stirred in an oil bath at 45 °C for 15 h (TLC monitored) until the reaction was complete. The reaction vessel was cooled to room temperature and the mixture quenched with water or a saturated solution of  $NH_4Cl$  (20 cm<sup>3</sup>). The organic layer was diluted with CH<sub>2</sub>Cl<sub>2</sub> and washed with a saturated solution of NH<sub>4</sub>Cl ( $3 \times 30$  cm<sup>3</sup>). The combined aqueous layers were extracted with  $CH_2Cl_2$  (3 × 30 cm<sup>3</sup>), dried over anhydrous MgSO<sub>4</sub>, and the solvent removed in vacuo. The crude products were then purified by twice repeated column chromatography on silica gel using CH<sub>2</sub>Cl<sub>2</sub> as eluent. The pure products were obtained as yellow needles.

*Method B.* In comparison with method A 13.5 mg palladium diacetate (0.06 mmol) was used instead of 20 mg palladium di(triphenylphosphine) dichloride (0.06 mmol) and 20 cm<sup>3</sup> toluene and 3 cm<sup>3</sup> DMF replaced 20 cm<sup>3</sup> THF.

*Method C.* In comparison with method A 43 mg bis(dibenzylideneacetone)palladium(0) (0.07 mmol) was used instead of 20 mg palladium di(triphenylphosphine) dichloride (0.06 mmol) and 1 cm<sup>3</sup> DIEA (6 mmol) replaced 0.83 cm<sup>3</sup> triethylamine (6 mmol).

Dimethyl 2-[[4-[3,5,7-tris[4-(ethoxycarbonyl)phenyl]-1adamantyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**17a**)

Method A: yield 25%; method B: yield 31%; method C: yield 54%; analytical data were identical with those published in Ref. [67].

Dimethyl 2-[[4-[[4-[3,5,7-tris[4-(ethoxycarbonyl)phenyl]-1-adamantyl]phenyl]ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (**17b**,  $C_{76}H_{64}N_2O_{10}$ )

Method A: yield 28%, m.p.: 164 °C; method B: yield 57%, m.p.: 162 °C; method C: yield 52%, m.p.: 163 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.49-8.52$  (d, J = 8.00 Hz, 1H, CH arom.), 8.22-8.42 (d, J = 8.72 Hz, 6H, CH arom.), 8.08-7.12 (d, J = 8.00 Hz, 1H, CH arom.), 7.72-7.75 (d, J = 7.20 Hz, 1H, CH arom.), 7.53–7.54 (dd, J = 7.20 Hz, J = 7.60 Hz, 4H, CH arom.), 7.45-7.49 (d, 8H, J = 8.72 Hz, CH arom.), 7.30–7.35 (d, 2H, J = 8.69 Hz), 7.10–7.14 (d, 2H, J = 8.60 Hz, CH arom.), 7.01–7.10 (m, 2H, CH arom.), 6.84–6.89 (dd, J = 1.76 Hz, J = 1.76 Hz, 1H, 6'-CH), 5.69-5.76 (m, 1H, 7'-CH), 5.34-5.67 (t, J = 2.2 Hz, 1H, 8'-CH), 4.90–5.94 (dt, J = 8.00 Hz, J = 2.00 Hz, 8'a-CH), 4.53–4.57 (q, 6H, J = 7.12 Hz, 3CH<sub>2</sub> ethyl ester), 3.95 (s, 3H, 3'-CH<sub>3</sub>), 3.41 (s, 3H, 2'-CH<sub>3</sub>), 2.14 (two overlapping s, 12H), 1.41 (t, 9H, J = 7.12 Hz, 3CH<sub>3</sub> ethyl ester) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 167.46$  (3'-CO), 165.22 (2'-CO), 163.15 (CO ethyl ester), 154.06, 149.68, 148.63, 148.60, 146.98, 142.78, 141.58, 138.97, 138.46, 138.12, 136.85, 136.67, 133.37, 133.25, 131.70, 130.13, 129.82, 128.79, 127.43, 124.79, 124.36, 123.75, 123.36, 122.07, 121.64, 121.38, 118.07, 116.79, 113.63, 109.83, 94.43 (acetylenic C), 93.45 (acetylenic C), 89.76 (acetylenic C), 89.34 (acetylenic C), 88.36, 64.63 (8'a-C), 63.20 (spiro-C), 61.51, 52.36 (3'-CH<sub>3</sub>), 51.46 (2'-CH<sub>3</sub>), 46.48, 39.65, 30.36, 26.20, 24.67, 23.38, 22.84, 22.10, 14.41, 12.12 ppm; IR (KBr):  $\bar{v} = 3,098-3,032$  (C–H, arom.), 2,895-2,980 (C-H, aliph.), 2,223 (acetylenic bond), 1,746 (3'-C=O), 1,706 (CO ester), 1,646 (2'-C=O), 1,584 (C=N), 1,451 (C=C), 1,346, 1,252, 1,181, 1,127, 968, 891, 766 cm<sup>-1</sup>; MS: m/z = 1,164.46 [M<sup>+</sup>].

 $(17c,\,C_{84}H_{68}N_2O_{10})$ 

Method A: yield 27%, m.p.: 143 °C; method B: yield 65%, m.p.: 141 °C; method C: yield 50%, m.p.: 144 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.56-8.59$  (d, J = 8.00 Hz, 1H, CH arom.), 8.32–8.40 (d, J = 8.72 Hz, 6H, CH arom.), 8.11–7.15 (d, J = 8.00 Hz, 1H, CH arom.), 7.70–7.74 (d, J = 7.20 Hz, 1H, CH arom.), 7.52–7.55 (dd, J = 7.20 Hz,

J = 7.60 Hz, 6H, CH arom.), 7.44–7.48 (d, 10H, J = 8.72 Hz, CH arom.), 7.33–7.36 (d, 2H, J = 8.69 Hz), 7.15–7.19 (d, 2H, J = 8.60 Hz, CH arom.), 7.12–7.16 (m, 2H, CH arom.), 6.80–6.86 (dd, J = 1.76 Hz, J = 1.76 Hz, 1H, 6'-CH), 5.76–5.84 (m, 1H, 7'-CH), 5.33–5.37 (t, J = 2.2 Hz, 1H, 8'-CH), 4.96–5.99 (dt, J = 8.00 Hz, J = 2.00 Hz, 8'a-CH), 4.56–4.60 (q, 6H, J = 7.12 Hz, 3CH<sub>2</sub> ethyl ester), 3.91 (s, 3H, 3'-CH<sub>3</sub>), 3.43 (s, 3H, 2'-CH<sub>3</sub>), 2.13 (two overlapping s, 12H), 1.45 (t, 9H, J = 7.12 Hz, 3CH<sub>3</sub> ethyl ester) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 166.99$  (3'-CO), 164.36 (2'-CO), 162.98 (CO ethyl ester), 153.16, 149.48, 148.37, 148.56, 146.85, 142.69, 141.32, 138.87, 138.36, 138.10, 136.71, 136.63, 133.37, 133.54, 131.36, 130.28, 129.87, 128.71, 127.40, 124.96, 124.37, 123.84, 123.30, 122.11, 121.87, 121.29, 118.64, 116.73, 113.64, 109.52, 94.39 (acetylenic C), 93.43 (acetylenic C), 89.76 (acetylenic C), 89.32 (acetylenic C), 88.26, 64.64 (8'a-C), 63.21 (spiro-C), 61.54, 52.34 (3'-CH<sub>3</sub>), 51.48 (2'-CH<sub>3</sub>), 46.47, 39.63, 30.48, 26.21, 24.60, 23.76, 22.81, 22.13, 14.54, 12.17 ppm; IR (KBr):  $\bar{\nu} = 3,089-3,019$  (C–H, arom.), 2,890-2,979 (C-H, aliph.), 2,219 (acetylenic bond), 1,743 (3'-C=O), 1,708 (CO ester), 1,643 (2'-C=O), 1,587 (C=N), 1,449 (C=C), 1,343, 1,252, 1,180, 1,132, 967, 895, 760 cm<sup>-1</sup>; MS: m/z = 1,264.49 [M<sup>+</sup>].

Dimethyl (4'aS)-2',4'a-dimethyl-2-[[4-[3,5,7-tris[4-(ethoxycarbonyl)phenyl]-1-adamantyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate (17d)

Method A: yield 22%; method B: yield 34%; method C: yield 52%; analytical data identical with those published in Ref. [67].

Dimethyl (4'aS)-2',4'a-dimethyl-2-[[4-[[4-[3,5,7-tris[4-(ethoxycarbonyl)phenyl]-1-adamantyl]phenyl]ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo-[1,2-b]pyridazine]-6',7'-dicarboxylate

(17e, C<sub>78</sub>H<sub>68</sub>N<sub>2</sub>O<sub>10</sub>)

Method A: yield 24%, m.p.: 134 °C; method B: yield 68%, m.p.: 133 °C; method C: yield 49%, m.p.: 134 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.37 - 8.39$  (d, J = 8.04 Hz, 1H, CH arom.), 8.11-8.19 (d, 6H, J = 8.72 Hz), 8.06-8.09(d, J = 8.00 Hz, 1H, CH arom.), 7.78–7.81 (d. J = 7.20 Hz, 1H, CH arom.), 7.65-7.66 (dd, J = 7.20 Hz, J = 7.60 Hz, 4H, CH arom.), 7.55–7.61 (d, 8H, J = 8.72 Hz, CH arom.), 7.42–7.45 (d, 2H, J = 8.60 Hz), 7.11–7.17 (d, 2H, J = 8.60 Hz), 7.00–7.08 (m, 2H, CH arom.), 5.69-5.73 (m, 1H, 7'-CH), 5.13-5.15 (dt, J = 8.00 Hz, J = 2.00 Hz, 8'-CH), 4.53-4.56 (q, 6H)J = 7.12 Hz, 3CH<sub>2</sub> ethyl ester), 3.95 (s, 3H, 3'-CH<sub>3</sub>), 3.51 (s, 3H, 2'-CH<sub>3</sub>), 2.17 (two overlapping s, 12H), 2.13 (s, 3H, 6'-CH<sub>3</sub>), 1.60 (s, 3H, 8'-CH<sub>3</sub>), 1.45 (t, 9H, J = 7.12 Hz, 3CH<sub>3</sub> ethyl ester) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 167.62 (3'-CO), 165.42 (2'-CO), 163.36 (CO ethyl ester), 152.85, 150.33, 148.72, 148.78, 147.54, 142.56, 142.30, 139.68, 138.63, 138.67, 136.14, 136.81, 133.66, 133.47, 131.26, 130.09, 129.79, 128.76, 127.63, 124.46, 124.19, 124.49, 123.36, 122.28, 121.79, 121.53, 118.39, 117.47, 112.49, 105.80, 94.89 (acetylenic C), 93.59 (acetylenic C), 89.98 (acetylenic C), 89.62 (acetylenic C), 88.38, 65.67 (8'a-C), 63.45 (spiro-C), 61.81, 52.46 (3'-CH<sub>3</sub>), 51.27 (2'-CH<sub>3</sub>), 46.95, 39.64, 30.20, 26.37, 24.57, 23.60, 22.23, 22.74 (6'-CH<sub>3</sub>), 22.44, 21.16 (8'-CH<sub>3</sub>), 14.16, 12.62 ppm; IR (KBr):  $\bar{\nu} = 3,079-3,006$  (C–H, arom.), 2,893–2,967 (C–H, aliph.), 2,225 (acetylenic bond), 1,749 (3'-C=O), 1,706 (CO ester), 1,685 (2'-C=O), 1,583 (C=N), 1,454 (C=C), 1,382, 1,273, 1,175, 1,136, 957, 886, 747 cm<sup>-1</sup>; MS: *m/z* = 1,192.49 [M<sup>+</sup>].

Dimethyl (4'aS)-2',4'a-dimethyl-2-[[4-[[4-[[4-[[4-[3,5,7-tris-[4-(ethoxycarbonyl)phenyl]-1-adamantyl]phenyl]ethynyl]phenyl]ethynyl]phenyl]ethynyl]spiro[9H-fluorene-9,5'(4'aH)-pyrrolo[1,2-b]pyridazine]-6',7'-dicarboxylate

 $(17f, C_{86}H_{72}N_2O_{10})$ 

Method A: yield 22%, m.p.: 121 °C; method B: yield 62%, m.p.: 119 °C; method C: yield 45%, m.p.: 119 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 8.49-8.53$  (d, J = 8.04 Hz, 1H, CH arom.), 8.25-8.29 (d, 6H, J = 8.72 Hz), 8.17-8.22 (d, J = 8.00 Hz, 1H, CH arom.), 7.84–7.86 (d, J = 7.20 Hz, 1H, CH arom.), 7.64–7.68 (dd, J = 7.20 Hz, J = 7.60 Hz, 6H, CH arom.), 7.50-7.56 (d, 8H, J = 8.72 Hz, CH arom.), 7.45–7.49 (d, 2H, J = 8.60 Hz), 7.10–7.16 (d, 2H, J = 8.60 Hz), 7.04–7.07 (m, 2H, CH arom.), 5.70–5.75 (m, 1H, 7'-CH), 5.06–5.09 (dt, J = 8.00 Hz, J = 2.00 Hz, 8'-CH), 4.50–4.53 (q, 6H, J = 7.12 Hz, 3CH<sub>2</sub> ethyl ester), 3.91 (s, 3H, 3'-CH<sub>3</sub>), 3.46 (s, 3H, 2'-CH<sub>3</sub>), 2.19 (two overlapping s, 12H), 2.11 (s, 3H, 6'-CH<sub>3</sub>), 1.64 (s, 3H, 8'-CH<sub>3</sub>), 1.44 (t, 9H, J = 7.12 Hz, 3CH<sub>3</sub> ethyl ester) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 167.64$  (3'-CO), 165.40 (2'-CO), 163.36 (CO ethyl ester), 152.76, 150.29, 148.64, 148.76, 147.50, 142.47, 142.29, 139.76, 138.65, 138.58, 136.09, 136.76, 133.63, 133.45, 131.27, 130.03, 129.84, 128.87, 127.60, 124.46, 124.23, 124.54, 123.38, 122.46, 121.85, 121.46, 118.76, 117.28, 112.46, 105.76, 94.68 (acetylenic C), 93.43 (acetylenic C), 89.82 (acetylenic C), 89.60 (acetylenic C), 88.36, 65.46 (8'a-C), 63.67 (spiro-C), 61.78, 52.40 (3'-CH<sub>3</sub>), 51.36 (2'-CH<sub>3</sub>), 46.84, 39.60, 30.28, 26.45, 24.50, 23.64, 22.19, 22.69 (6'-CH<sub>3</sub>), 22.40, 21.17 (8'-CH<sub>3</sub>), 14.16, 12.46 ppm; IR (KBr):  $\bar{\nu} = 3,088-3,022$  (C–H, arom.), 2,890–2,961 (C–H, aliph.), 2,221 (acetylenic bond), 1,745 (3'-C=O), 1,703 (CO ester), 1,682 (2'-C=O), 1,583 (C=N), 1,451 (C=C), 1,388, 1,270, 1,167, 1,146, 958, 882, 743 cm<sup>-1</sup>; MS: m/z = 1,292.52 [M<sup>+</sup>].

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#### References

- 1. Bouas-Laurent H, Dürr H (2001) Pure Appl Chem 73:639
- Irie M (2001) In: Feringa BL (ed) Molecular switches. Wiley-VCH, Weinheim
- Mizokuro T, Mochizuki H, Kobayashi A, Horiuchi S, Yamamoto N, Tanigaki N (2004) Chem Mater 16:3469
- Mizokuro T, Mochizuki H, Xiaoliang M, Horiuchi S, Tanaka N, Tanigaki N (2003) Jpn J Appl Phys 42:983
- 5. Hirshberg Y, Fischer E (1953) J Chem Soc 629
- Persano L, Mele E, Athanassiou A, Cingolani R, Psignano D (2006) Chem Mater 18:4171
- 7. Kanakkanatt SV (1997) Proc SPIE Int Soc Opt Eng 3227:218
- 8. Berkovic G, Krongauz V (2000) Chem Rev 100:1741
- 9. Uznanski P (2003) Langmuir 19:1919
- 10. Guglielmetti R (1990) In: Dürr H, Bouas-Laurent H (eds) Photochromism: molecules and systems. Elsevier, Amsterdam
- 11. Ahmed SA, Hartmann T, Dürr H (2008) J Photochem Photobiol 200:50
- 12. Dürr H, Bouas-Laurent H (eds) (1990, 2003) In: Photochromism: molecules and systems. Elsevier, Amsterdam
- Dürr H (1995) In: Horspool WM, Song PS (eds) Organic photochemistry and photobiology. CRC Press, Boca Raton
- Dürr H (1999) In: Crano JC, Guglielmetti RJ (eds) Organic photochromic and thermochromic compounds. Plenum Press, New York
- 15. Dürr H (1989) Angew Chem Int Ed 28:413
- Bleisinger H, Scheidhauer P, Dürr H, Wintgens V, Valat P, Kossanyi J (1998) J Org Chem 63:990
- 17. Gross H, Dürr H (1982) Angew Chem Int Ed 21:216
- Terazono Y, Kodis G, Andreasson J, Jeong G, Brune A, Hartmann T, Dürr H, Moore LA, Moore SA, Gust G (2004) J Phys Chem B 108:1812
- Kodis G, Liddell PA, de la Garza L, Clausen PC, Lindsey JS, Moore LA, Moore SA, Gust D (2002) J Phys Chem A 106:2036
- 20. Weber C, Rustemeyer F, Dürr H (1998) Adv Mater 10:1348
- Ahmed SA, Hartmann T, Huch V, Dürr H, Abdel-Wahab AA (2000) J Phys Org Chem 13:539
- Tan YS, Ahmed SA, Dürr H, Huch V, Abdel-Wahab AA (2001) Chem Comm 14:1246
- 23. Ahmed SA (2005) Mol Cryst Liq Cryst 430:295
- 24. Ahmed SA, Dürr H (2005) Mol Cryst Liq Cryst 431:275
- 25. Ahmed SA (2004) Monatsh Chem 135:1173
- Ahmed SA, Abdel-Wahab AA, Dürr H (2003) J Photochem Photobiol 154:131
- 27. Ahmed SA (2002) J Phys Org Chem 15:392
- 28. Ahmed SA (2006) J Phys Org Chem 19:402
- 29. Ahmed SA (2007) J Phys Org Chem 20:564
- Fromm R, Ahmed SA, Hartmann T, Huch V, Abdel-Wahab AA, Dürr H (2001) Eur J Org Chem 21:4077
- 31. Ahmed SA, Pozzo JL (2008) J Photochem Photobiol 200:57
- 32. Dürr H, Schommer C, Münzmay T (1986) Angew Chem 25:565
- Dürr H, Thome A, Kilburg K, Bossmann S, Blasius E, Janzen K, Kranz C (1992) J Phys Org Chem 5:689
- 34. Dürr H (1994) Chimica 514
- 35. Dürr H, Amlung M, Rustemeyer F, Tan YS (1998) Deutsche Offenlegungs Schrift Pat 198 349 408

62. Marsden JA, Miller JJ, Shirtcliffe LD, Haley MM (2005) J Am Chem Soc 127:2464

from molecules to materials. Wiley-VCH, Weinheim

61. Nielsen MB, Diederich F (2005) Chem Rev 105:1837

60. Bunz UHF, Rubin Y, Tobe Y (1999) Chem Soc Rev 28:107

- 63. Bowling NP, Halter RJ, Hodges JA, Seburg RA, Thomas PS, Simmons CS, Stanton JF, McMahon RJ (2006) J Am Chem Soc 128:3291
- 64. Cornil J, Beljonne D, Calbert JP, Bredas (2001) Adv Mater 13:1053
- 65. Ahmed SA (2009) Tetrahedron 65:1373
- 66. Ahmed SA (2008) Res Lett Org Chem. Article ID 959372
- 67. Ahmed SA, Al-Raqa SY (2009) J Phys Org Chem. (in press)
- 68. Philip J, Martin A, Rhian T, Sharon M, Reszka P, Wood A, Lloyd R (1999) J Med Chem 42:2679
- 69. Gonzalo Rodriguez J, Luis Tejedor TPJ, Diaz C (2006) Tetrahedron 62:3355
- 70. Thyagarajan S, Liu A, Famoyin OA, Lamberto M, Galoppini E (2007) Tetrahedron 63:7550
- 71. Galoppini E, Guo W, Zhang W, Hoertz PG, Qu P, Meyer GJ (2002) J Am Chem Soc 124:7801
- 72. Galoppini E (2004) Coord Chem Rev 248:1283
- 73. Wei Q, Galoppini E (2004) Tetrahedron 60:8497
- 74. Guo W, Galoppini E, Rydja GI, Pardi G (2000) Tetrahedron Lett 41:7419
- 75. Galoppini E, Guo W, Qu P, Meyer GJ (2001) J Am Chem Soc 123:4342
- 76. Galoppini E, Guo W, Zhang W, Hoertz PG, Qu P, Meyer GJ (2002) J Am Chem Soc 124:7801
- 77. Piotrowiak P, Galoppini E, Wei Q, Meyer GJ, Wiewior P (2003) J Am Chem Soc 125:5278
- 78. Wang D (2004) PhD thesis. Rutgers University
- 79. Wang D, Schlegel JM, Galoppini E (2002) Tetrahedron 58:6027
- 80. Hoertz PG, Carlisle RA, Meyer GJ, Wang D, Piotrowiak P, Galoppini E (2003) Nano Lett 3:325
- 81. Zarwell S, Rück-Braun K (2008) Tetrahedron Lett 49:4020
- 82. Gautron R (1968) Bull Soc Chim France 3190
- 83. Schönberg A (1958) In: Präparative Organische Photochemie. Springer, Berlin Heidelberg New York

- 37. Ahmed SA, Abdel-Wahab AA, Dürr H (2003) In: Horspool WM, Lenci F (eds) CRC handbook of organic photochemistry and photobiology, 2nd edn. CRC Press, New York
- 38. Dürr H (1989) Angew Chem 101:427
- 39. Dürr H (1984) Zeitschr TH Leuna-Merseburg 26:664
- 40. Dürr H, Gross H, Zils KD (1983) Deutsche Offenlegungs Schrift Pat 32 20 275 A1
- 41. Dürr H, Jönsson HP, Scheidhauer P, Münzmay T, Spang P (1985) Deutsche Offenlegungs Schrift Pat 3521432 5
- 42. Dürr H, Janzen KP, Thome A, Braun B (1988) Deutsche Offenlegungs Schrift Pat 3521432 5
- 43. Dürr H, Gross H, Zils KD, Hauck G, Hermann H (1983) Chem Ber 116:3915
- 44. Dürr H, Spang P (1984) Deutsche Offenlegungs Schrift Pat 32 20 2571
- 45. Ahmed SA (2000) PhD thesis. Saarland-Assiut Universities
- 46. Dürr H, Jeonsson HP, Scheidhauer P, Münzmay T, Spang P (1985) Deutsche Offenlegungs Schrift Pat 35214325
- 47. Burtscher P, Dürr H, Rheinberger V, Salz U (1995) German Pat 195200160
- 48. Service RF (2001) Science 294:2442
- 49. Rodriguez JG, Tejedor JL, La Parra T, Diaz C (2006) Tetrahedron 62:2355
- 50. Price DW, Tour JM (2003) Tetrahedron 59:3131
- 51. Montemerlo MS, Love GC, Opiteck GJ, Goldhaber-Gordon D, Ellenbogen JC (1996) Technologies and designs for electronic nanocomputers. MITRE Corporation
- 52. Tour JM (2000) Acc Chem Res 33:791
- 53. Tour JM, Kozaki M, Seminario J (1998) J Am Chem Soc 120:8486
- 54. Zhou C, Deshpande MR, Reed MA, Jones L, Tour JM (1997) Appl Phys Lett 71:611
- 55. Chen J, Reed MA, Rawlett MA, Tour JM (1999) Science 1999:1550
- 56. Chen J, Wang W, Reed MA, Rawlett AM, Price DW, Tour JM (2000) Appl Phys Lett 77:1224
- 57. Chen J, Wang W, Reed MA, Rawlett AM, Price DW, Tour JM (2001) Mater Res Soc Symp Proc 5582:H321
- 58. Diederich F, Stang PJ, Tykwinski RR (2005) In: Acetylene chemistry: chemistry, biology and material science. Wiley-VCH, Weinheim